



COMPUTATIONAL ANDSTRUCTURAL BIOTECHNOLOGY

JOURNAL



journal homepage: www.elsevier.com/locate/csbj

MIAOME: Human microbiome affect the host epigenome

Lidan Wang^{a,1}, Wei Zhang^{b,1}, Xianglu Wu^{c,1}, Xiao Liang^a, Lijie Cao^a, Jincheng Zhai^a, Yiyang Yang^a, Qiuxiao Chen^a, Hongqing Liu^a, Jun Zhang^a, Yubin Ding^{c,*}, Feng Zhu^{b,*}, Jing Tang^{a,c,*}

^a School of Basic Medicine, Chongqing Medical University, Chongqing 400016, China

^b College of Pharmaceutical Sciences, Zhejiang University, Hangzhou 310058, China

^c Joint International Research Laboratory of Reproductive and Development, Department of Reproductive Biology, School of Public Health, Chongqing Medical University, Chongqing 400016, China

81.8

ARTICLE INFO

Article history: Received 24 February 2022 Received in revised form 11 May 2022 Accepted 12 May 2022 Available online 17 May 2022

Keywords: Human microbiome Host epigenome Epigenetic modification Microbiota-derived metabolites Microbiota-secreted proteins

ABSTRACT

Besides the genetic factors having tremendous influences on the regulations of the epigenome, the microenvironmental factors have recently gained extensive attention for their roles in affecting the host epigenome. There are three major types of microenvironmental factors: microbiota-derived metabolites (MDM), microbiota-derived components (MDC) and microbiota-secreted proteins (MSP). These factors can regulate host physiology by modifying host gene expression through the three highly interconnected epigenetic mechanisms (e.g. histone modifications, DNA modifications, and non-coding RNAs). However, no database was available to provide the comprehensive factors of these types. Herein, a database entitled 'Human Microbiome Affect The Host Epigenome (MIAOME)' was constructed. Based on the types of epigenetic modifications confirmed in the literature review, the MIAOME database captures 1068 (63 genus, 281 species, 707 strains, etc.) human microbes, 91 unique microbiota-derived metabolites & components (16 fatty acids, 10 bile acids, 10 phenolic compounds, 10 vitamins, 9 tryptophan metabolites, etc.) derived from 967 microbes; 50 microbes that secreted 40 proteins; 98 microbes that directly influence the host epigenetic modification, and provides 3 classifications of the epigenome, including (1) 4 types of DNA modifications, (2) 20 histone modifications and (3) 490 ncRNAs regulations, involved in 160 human diseases. All in all, MIAOME has compiled the information on the microenvironmental factors influence host epigenome through the scientific literature and biochemical databases, and allows the collective considerations among the different types of factors. It can be freely assessed without login requirement by all users at: http://miaome.idrblab.net/ttd/

© 2022 The Author(s). Published by Elsevier B.V. on behalf of Research Network of Computational and Structural Biotechnology. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Epigenome plays an essential role in regulating countless biological processes in kinds of developmental and environmental contexts [1,2]. The regulations of the epigenome are dynamic [3,4] and frequently influenced by multiple factors including genetic, physiological, and environmental factors (*e.g.* diet, lifestyle) [5–7], and these influences conduce to growth, development, and disease risk [8]. Besides the above factors having tremendous impacts on the epigenome, the microenvironmental factors have recently gained extensive attention for their roles in affecting the host epigenome [9–11]. Particularly, the human microbiome, which provides microenvironmental factors, can trigger host epigenetic modification [12]. Increasing evidence shows that the microbes, which as one type of environment hint, can regulate host physiology by modifying host gene expression through three highly interconnected epigenetic mechanisms (*e.g.* histone modification, DNA modification, and non-coding RNAs) [13–15].

There are three major types of microenvironmental factors that are continually considered in current epigenome studies [16–20]: (i) *microbiota-derived metabolites* (MDM, including fatty acids, bile acids, phenolic compounds, vitamins, tryptophan metabolites,

https://doi.org/10.1016/j.csbj.2022.05.024

Abbreviations: MDM, microbiota-derived metabolites; MDC, microbiota-derived components; MSP, microbiota-secreted proteins; ncRNAs, non-coding RNAs; LPS, lipopolysaccharide; HDACs, histone deacetylases; miRNA, microRNA; EM, Epigenetic molecule.

^{*} Corresponding authors at: School of Basic Medicine, Chongqing Medical University, Chongqing 400016, China (J. Tang).

E-mail addresses: dingyb@cqmu.edu.cn (Y. Ding), zhufeng@zju.edu.cn (F. Zhu), tang_jing@cqmu.edu.cn (J. Tang).

¹ The authors wish it to be known that, in their opinion, the first three authors should be regarded as joint First Authors.

^{2001-0370/© 2022} The Author(s). Published by Elsevier B.V. on behalf of Research Network of Computational and Structural Biotechnology. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

amino acids, choline metabolites, polyamines, phenolic compounds, etc.) are crucial in mediating host epigenetic reprogramming [21], affecting epigenetic mechanisms that contribute to human diseases [22], and revealing specific biomarkers and potential therapeutic targets [23]; (ii) microbiota-secreted proteins (MSP, including Listeriolysin O, HP0175, NuE, RomA, etc.) are tools used by pathogenic bacteria to induce epigenetic changes in the host to favor infection [24], they can alter the epigenetic type and gene expression pattern in the host cells [25], and facilitate human disease initiation and progression [26-28]; (iii) microbiota-derived components (MDC, including lipopolysaccharide, flagellum, βglucan, etc.), for example, lipopolysaccharide (LPS) from Fusobacterium nucleatum induced transient acetylation of histone H3 lysine 9 in oral epithelial cells through the pattern recognition receptor pathway [29]. Besides, there are indications that the altered microbiome composition or decreased/increased abundances of microbes also have impacts on the host epigenetic modifications [30–32]. Since these microbe-induced epigenetic modifications are necessary for proper homeostasis in the host [14], the accumulation of anthropogenic factors that define such modifications could provide important insights into the current understanding of the microbiome and the study of its interaction with host epigenetic mechanisms.

Up to now, a number of human epigenome-related databases have been constructed and are currently open, the most of which focus on providing epigenetic modifications data related to diseases (CFEA [33], dbEM [34], DANCER [35], EWASdb [36], DNMIVD [37], EWAS Data Hub [38], MethCNA [39], MOBCdb [40], MethylomeDB [41], DiseaseMeth [42]), while some others are specialized in containing particular classes of DNA modifications (EWAS Atlas [43], REBASE [44], MethDB [45], DNAmod [46] and MethMotif [47]), or histone modifications (HHMD [48], dbHiMo [49]). However, there still has no database has been developed for providing the data on microenvironment factors that affect the host epigenetic modifications.

Here, a newly developed database, as illustrated in Fig. 1, the human microbiome affect the host epigenome (MIAOME) was therefore introduced. First, a comprehensive literature review on host epigenome was conducted. Different types of epigenetic modifications involved in 20 histone modifications, 4 types of DNA modifications, and 490 types of ncRNAs regulations, were included in MIAOME. Second, based on these various epigenetic modifications, three microenvironmental factors (MDM, MDC and MSP) modulating each host epigenome were collected via literature review. Third, the crosslinks of each epigenome/microbe to disease associations were systematically discovered. These crosslinks could facilitate understanding the pathogenesis of diseases by integrating microbiome and epigenome. In a word, the MIAOME comprehensively provided the MDM, MDC and MSP information that affects the host epigenome, which allows the collective considerations among the different types of factors (among MDM, MDC and MSP; between microbiota and host). As the epigenetic modifications are affected by the microenvironmental factors [9–11], the MIAOME is expected to have implications for the future practice of epigenome-related studies on human physiology and health [23,50].

2. Methods

2.1. Epigenetic modifications data collection.

Epigenetic modification refers to stable and heritable changes in gene expression and cell function without altering its DNA sequence, usually caused by genetic and environmental factors. In the host, the epigenetic mechanism can regulate gene expression through a range of reversible epigenetic modifications [51], such as histone acetylation, histone phosphorylation, histone ubiquitylation, DNA methylation, and non-coding RNAs. Because of the diverse types of epigenetic modifications, the data of specific types of epigenetic modifications were collected by literature reviews in PubMed [52] using the keyword combinations of: 'Epigenetic modification', 'Epigenetic mechanism', 'histone + modification', 'DNA modification', *etc.* All retrieved information on epigenetic modification is the basis of earlier published literature (until December 31, 2021). The reference literature was reviewed to ascertain the diverse types of epigenetic modification. Based on a particular type of modification, we systematically collected the influence of microenvironmental factors on the host epigenome in the subsequent procedure.

2.2. The epigenetic modifications-associated microenvironmental factors collection.

Based on a particular type of modification, we systematically collected the influence of microenvironmental factors including MDM, MDC and MSP on the host epigenome in this procedure. In particular, the human microbe is well-known to produce varieties of metabolites, such as short-chain fatty acids including acetic acid, butyric acid, and propionic acid that can regulate a range of host physiological functions [53,54]. These MDMs act as critical modulators of epigenetic enzymes, influencing the DNA methylation and histone modifications, and mediating changes in microRNA expression [55,56]. And, the components of microbe cells (e.g. lipopolysaccharide of the cell wall, peptidoglycan of the cell wall, flagellin of flagellum) have been shown to influence the epigenetic modifications in host cells [57,58], and then induce changes in cellular expression profiles [17]. Besides, recent studies indicated that the microbiota-secreted proteins (such as Ankyrin A, Listeriolysin O, Rv2966c, etc.) were essential in reprogramming host epigenome [25], alternating the host cell epigenotype [59–61], and inducing the DNA methylation in host cells [50,62]. These factors are critical for further understanding the mechanisms of microbial infection [63] and discovering the epigenetic therapy potential targets [25].

PubMed [52] was systematically searched to find the impacts of host epigenome by MDM, MDC, and MSP. Particularly, keyword combinations such as 'human microbe + 'epigenetic modification", 'human microbe metabolite + 'epigenetic modification", 'human microbe metabolite + 'epigenetic mechanism", 'human microbiota metabolite + 'DNA modification", 'human microbiota metabolite + 'histone modification", 'microbe protein + 'epigenetic modification", 'human microbe protein + 'ncRNA", and 'human microbe protein + 'histone modification" were adopted for literature reviews. All retrieved information on microbial factors that influenced host epigenome is the basis of earlier published literature (until December 31, 2021). The reference literature was reviewed to ascertain the association of microenvironmental factors to each type of epigenetic modification. These MDM, MDC, and MSP, whose associations with host epigenome could not be confirmed based on the reference, were discarded. As a result, the collected information included the names of microbes, the specific types of MDM, MDC, and MSP, various types of epigenetic modifications (such as DNA methylation, histone acetylation, non-coding RNA regulation), and the effects of these microbial factors on the host epigenome (promotion/inhibition of host DNA methylation, up/ down-regulation of host non-coding RNA, etc.).

Moreover, relevant information on the validated microbe/ MDM/MDC/MSP, such as taxonomy ID, taxonomic lineage, oxygen dependency/tolerance, Gram-staining classification, diverse synonyms of each microbe, disease-related information, metabolite formula/synonyms/external links and so on was retrieved from



Fig. 1. MIAOME comprehensively described microenvironmental factors affecting human microbes, which allowed the collective consideration among different types of factors (between microbiota-derived metabolites & components, and microbiota-secreted proteins). By providing the crosslinks to available databases (EWASdb [35], DNMIVD [36], gutMDisorder [65] and GIMICA [66]) with host phenotypes information, MIAOME further enabled the collective consideration between the microenvironmental factors and host phenotypes.

online databases such as NCBI PubMed, NCBI Taxonomy [64], GIMICA [65], gutMDisorder [66], VMH [67] and UniProt [68].

2.3. Epigenome molecular collection.

The epigenome is a complete description of all chemical modifications of DNA and histones, which regulate the expression of genes in the genome. The epigenome defines the unique gene expression pattern and resulting cellular behavior in different cell types [69]. The epigenome is highly cell-type-specific [70,71]. Therefore, it's crucial to collect information on the influence of microenvironmental factors on the epigenome in a specific cell of the host. Moreover, identification of the genomic location of epigenetic modifications will assist in elucidating its functions [72]. The epigenetic modifications at different genomic locations (e.g. gene body, promoter, exon, and intron) may have different impacts on regulating the expression of genes [73] and even can lead to an opposite genetic outcome [74]. As a result, we collected the critical information on epigenetic modifications influenced by the abovementioned three microbial factors, which include the cell types of host, biological conditions, and genomic locations of epigenome modifications. Besides, we have also provided the official full names, synonyms, UniProt entry, and Ensemble ID for these genes through searching Uniprot [68] and Ensembl [75]. In addition, according to the host gene data we have collected, the diverse types of diseases associated with these genes were also collected by further searching in the EWASdb [36] and DNMIVD [37] databases. All information can be retrieved using a variety of search strategies in both the homepage and subpages of the MIAOME.

2.4. Data standardization, access, and retrieval.

To facilitate access to MIAOME data convenient for all users, the collected information was carefully cleaned and systematically standardized. These standardizations included: (i) all microbes were standardized using NCBI Taxonomy [64], the extended data of each microbe could be accessed using crosslinks to NCBI Taxonomy [64], gutMDisorder [66], GIMICA [65], GOLD [76]; (ii) all MDM

were standardized using VMH [67] and HMDB [77], the extended data of each metabolite could be accessed using crosslinks to KEGG [78] and METLIN [79]; (iii) all MSP were standardized using Uni-Prot [68] database; (iv) all genes were standardized using Ensembl [75] database; (v) all diseases in MIAOME were standardized by the latest version of International Classification of Diseases (ICD-11, officially released by World Health Organization [80]) and the extended data of each gene could be accessed using crosslinks to UniProt [68], EWASdb [36] and DNMIVD [37].

3. Results and discussion

3.1. MIAOME database content.

The regulations of microbes on the host epigenome were frequently discovered to be caused by their derived metabolites, components, or secreted proteins [81,82]. For example, short-chain fatty acids produced by gut microbes promote the differentiation of naïve T cells into Treg by inhibiting histone deacetylases (HDACs), and this metabolite may cause abnormal immune response via epigenetic modifications, and lead to autoimmune diseases [23]. To gain a comprehensive understanding of the influence of microbes on the host epigenome, three microenvironmental factors were systematically reviewed and collected in MIAOME for each epigenetic modification. Consequently, the collected data included the names of microbes, MDM (16 fatty acids, 10 bile acids, 10 phenolic compounds, 11 vitamins, 9 tryptophan metabolites, 5 amino acids, 5 choline metabolites, etc.), MDC (lipopolysaccharide, flagellum, β-glucan, *etc.*), and MSP (HP0175, Listeriolysin O, NuE; RomA, Flagellin, Ankyrin A, InIB, etc.), various types of epigenetic modifications (DNA methylation, histone acetylation, histone phosphorylation, histone crotonylation, histone ubiquitination, histone methylation, miRNA regulations, etc.)

Furthermore, the genes modified by the above microenvironmental factors and the genome locations of modification were also collected, and provided 160 diseases related to them. Meanwhile, MIAOME provided the influences of each microbial factor on the host epigenome (promotion/inhibition of host DNA methylation, up/down-regulation of host non-coding RNA, *etc.*). All epigenetic modification-related data caused by microbial factors can be retrieved using various search strategies on the homepage and subpages of the MIAOME database as shown in Fig. 2.

Besides the microbial factors above discussed, a variety of other factors (including ethnicity, age, lifestyle, and environment) have been reported to influence the host epigenome [83]. For instance, maternal diabetes/obesity during pregnancy could mediate the altered infant methylation patterns in the cord blood [84]. The perinatal nicotine exposure remodeled the DNA methylation in off-spring spermatozoa [85]. Comprehensive considerations of microbial factors, host physiology, and environment could provide new insight into the host epigenome [86]. Current MIAOME focused on microbe influenced the host epigenome and lacked other critical environmental factors. The collective consideration of various

factors types has great implications to serve comprehensive human health in the future.

3.2. MIAOME data and statistics.

All in all, the current version of MIAOME incorporates 1068 (63 genus, 281 species, 707 strains, *etc.*) human microbes, of which 967 produced 91 metabolites & components (valerate, lactate, folate, hydrogen sulfide, tryptophan, spermine, hexanoate, caproate, *etc.*), 50 secreted 40 proteins (Rv3763, Rv1988, LMP-1, Pneumolysin, TRP32, *etc.*); 98 microbes directly influence the host epigenetic modifications. These microbial factors modulate 514 types of epigenetic modifications, including (1) 4 types of DNA modifications (DNA methylation, DNA hypermethylation, DNA Hypomethylation, DNA demethylation); (2) 20 histone modifications (histone



Fig. 2. Sample web interfaces. There are six major web pages in MIAOME: 'Home' page, 'Microbiota-Derived Metabolite & Component' page, 'Microbiota-Secreted Protein' page, 'Microbiota' page, 'Epigenetic Molecule' page, and 'Download' page.



Fig. 3. An UpSetR plot shows the intersection of three types of microbe sets, which included microbe-affected host DNA modification, microbe-affected histone modification, and microbe-affected ncRNAs regulation. The brown line marks the microbes shared by three types of epigenetic modification.

acetylation, histone biotinylation, histone crotonylation, histone methylation, histone phosphorylation, histone ubiquitination, *etc.*) and (3) 490 ncRNAs regulations. Besides, we have also provided more than 12,000 host genes that are regulated by epigenetic modifications, 129 cell types, and 160 diseases related to these genes. With the data contained in MIAOME, we performed a statistical

analysis on the distribution of epigenetic modification information caused by microbial factors. An UpSetR plot (Fig. 3) shows the intersections of microbes that influenced three types of epigenetic modifications. In general, histone modifications were influenced by most microbes, and 264 microbes can influence the host gene expression through the three epigenetic modifications.

Home	Microbiota-Derived Metabolite & Component	Microbiota-Secreted Protein	Microbiota	Epigenetic Molecule	Download
		Search for the Microbe S	pecies (MIC)		
Search for	Microbiota Entries: 🚱				
				Search	Reset
Examples: L	actobacillus acidophilus; Resveratrol; R	v2966c; EIF3A			
Search for © Tips	MIC Entries by MIC Name based on Ta : Please select the phylum name first, the	axonomic Phylum: ? en a list of MIC name under the	selected phylum will	be avaiable for selection.	
	Step 1: Please Select a Phylum Name)		~	
	Step 2: Please Select a MIC Name			Search ✓	Reset
Home	Microbiota-Derived Metabolite & Component	Microbiota-Secreted Protein	Microbiota	Epigenetic Molecule	Download
Search for	Microbial Metabolite & Component En	tries: 🕜		-	
				Search	Reset
Examples:	equol; Lactobacillus acidophilus; ESR1				
Search for © Tips	Microbial Metabolite & Component En : Please select the metabolite type first, t	tries based on Metabolite Typ hen a list of metabolite name un	e: 😯 der the selected type	e will be avaiable for selection.	
	Step 1: Please Select a Metabolite Ty	ре		×	
	Step 2: Please Select a Metabolite Na	ime		Search	Reset
	Microbiota-Derived	Microbiota-Secreted			
Home	Metabolite & Component	Protein	Microbiota	Epigenetic Molecule	Download
Search for	Microbiota-Secreted Protein Entries:	0			
				Search	Reset
Examples:	Rv2966c; Mycobacterium tuberculosis H	37Rv; CNTN5			
Search for © Tips	Microbiota-Secreted Protein Entries b :: Please select the microbiota first, then a	y the Name of Protein based of a list of protein name under the s	on Microbiota: 😧	avaiable for selection.	
	Step 1: Please Select a Microbiota			~	
	Chan O: Diagon Colont - Destain Norma			Search	Reset
	Step 2: Please Select a Protein Name	Misrobioto Coorotod		×	
Home	Microbiota-Derived Metabolite & Component	Protein	Microbiota	Epigenetic Molecule	Download
Search for	Microbiota Entries: 😢				
				Search	Reset
Examples:	AB58; MC1170; Gardnerella vaginalis				
Search for © Tips	MIC Entries by the Name of MIC Name : Please select the Body Site first, then a	e based on Body Site: 0 list of MIC Name under the sele	ected type will be ava	aiable for selection.	
	Step 1: Please Select a Body Site			×	Dent
	Step 2: Please Select a MIC Name			✓ Search	Reset
Home	Microbiota-Derived Metabolite & Component	Microbiota-Secreted Protein	Microbiota	Epigenetic Molecule	Download
Search for	Epigenetic Molecule Entries: 😧				
				Search	Reset
Examples: I	Lactobacillus acidophilus; Resveratrol; R	v2966c; HES1			
Search for Tips	Epigenetic Molecule Entries by the Na : Please select the ICD11 code first, ther	ame of Epigenetic Molecule ba a list of Epigenetic Molecule un	sed on ICD11 Code der the selected type	e will be avaiable for selection.	
	Step 1: Please Select an ICD11 Code			~	

Fig. 4. MIAOME offers different ways to search the entries, based on users' preferences. Users can find microbiota-inducted epigenetic modifications through several approaches, such as microbe names, MDM names, MDC names, MSP names, and Epigenetic molecule names in MIAOME.

3.3. MIAOME Search.

To construct a database with plentiful data, applying a good search algorithm with more convenient and efficient for users is very important. As illustrated in Fig. 4, MIAOME offers different ways to search the entries, based on users' preferences. Users can find microbiota-inducted epigenetic modifications by searching microbe names (*e.g. 'Lactobacillus acidophilus'*), MDM names (*e.g.* 'equol'), MSP names (*e.g. 'Rv*2966c'), and their ID numbers (*e.g.* 'MC1011', 'MT104', 'PT007') among the entire textual component of MIAOME. The query can be submitted by entering keywords into the main searching frame. To facilitate a more customized input query, the wild characters of "*" and "?" are also supported in MIAOME. Moreover, users can click the corresponding '?' button to learn the detailed search steps. Fig. 5 demonstrates an example of searching by microbiota-derived metabolite name.

Searched results will be displayed on a separate web page. The page will display the basic information of searched queries (as shown in Fig. 5A). By clicking the "Meta Info" hyperlink button, the detailed information page of this metabolite will be displayed, this page is divided into three parts, "General Information of Metabolite", "The epigenetic modification information of this metabolite", "The microbes that produce this metabolite" (as

shown in Fig. 5B). The general information of this metabolite includes its "Meta Name", "Unify name", "Synonym", "Formula", "Inchi Key" and "Description". If you want to know more about this metabolite, you can link to other related databases by clicking the hyperlinks in the "External Links" field. "The epigenetic modification information of this metabolite" includes the epigenetic modification information influenced by the searched metabolite, such as "Modification Type", "Molecule", "Cell/Tissue Type", and "Modified sites". Moreover, detailed information on each epigenetic molecule can be browsed on a new webpage by clicking on the specific "EM Info" (as shown in Fig. 5C). "The microbes that produce this metabolite" includes the microbe information that produce the searched metabolite, such as "Microbiota Name" and "Description". By clicking on the specific "MIC Info", the general information of this microbe will be displayed (as shown in Fig. 5D).

3.4. MIAOME download functionality.

MIAOME also provides functions for downloading all MIAOME data from various customized links. As Fig. 2 shows, MIAOME allows users to download the data of microbes, MDM, MDC, MSP, epigenetic molecules, and the epigenetic modifications induced by MDM, MDC, and MSP. All data can be readily downloaded by



Fig. 5. An example for data accessing in MIAOME. Here, four different searches were supported, by MDM names, MDC names, the microbe names, and the epigenetic molecule names. (A) An example illustrates how to explore the epigenetic modifications through MDM names (Resveratrol is selected). (B) Typical search result webpage using the MDM name: Resveratrol as an example. The detailed information page of Resveratrol was displayed. Details of the metabolite are divided into three parts: General Information of Metabolite, Epigenetic modification information of this metabolite, and the microbes producing this metabolite. (C) The epigenetic molecule webpage consists of three major sections showing detailed information: General Information of EM, Metabolite/Component(s) and Protein(s) associated with this EM. (D) The microbe webpage consists of two major sections showing detailed information: General Information of MIC and Full List of Metabolite(s) Produced by This MIC.

simply clicking the corresponding "Click to Save" button. These files can beneficial to researchers that engaged in epigenomerelated studies and provide basic data resources for discovering targets of epigenetic therapy.

4. Conclusions

MIAOME is a user-friendly and new freely available online resource that compiles information on the microenvironmental factors influence host epigenome through scientific literature and biochemical databases. These microenvironmental factors included the MDM, MDC, and MSP. Manually curated information on the type of epigenetic modification, the cell type/tissue, and the genomic location of modification along with supporting reference literature has been collated and included in MIAOME. Moreover, the links between the epigenome and host phenotypes are collectively provided in MIAOME. MIAOME is expected to significantly advance our understanding of the epigenetic regulation underlying human physiology and disease. MIAOME has been smoothly running for months and tested from various sites around the world and is freely assessable without login requirement by all users at: http://miaome.idrblab.net/ttd/.

Contributors

J.T. conceived the idea and supervised the work. L.W., W.Z. performed the research. F.Z., W.Z. constructed the database, developed the software and wrote the scripts. L.W., X.W., X.L., L.C., J.Z., Y.Y., Q. C., H.L., J.Z., and Y.D. prepared the data. J.T. and L.W. wrote manuscript. All authors reviewed and approved the final version of the manuscript.

CRediT authorship contribution statement

Lidan Wang: Data curation, Writing - Original Draft, Writing -Review & Editing. Wei Zhang: Data curation, Software. Xianglu Wu: Data curation, Investigation. Xiao Liang: Visualization, Software. Lijie Cao: Visualization, Data curation. Jincheng Zhai: Visualization, Data curation. Yiyang Yang: Visualization, Data curation. Qiuxiao Chen: Visualization, Data curation. Hongqing Liu: Visualization, Data curation. Jun Zhang: Visualization, Data curation. Yubin Ding: Supervision, Funding acquisition. Feng Zhu: Conceptualization, Project administration. Jing Tang: Conceptualization, Writing - Original Draft, Writing - Review & Editing, Supervision, Funding acquisition.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This work was supported by the National Key Research and Development Program of China (2018YFC1004401) and the Science and Technology Research Program of Chongqing Municipal Education Commission (KJQN202100421).

References

 Dawson MA, Kouzarides T. Cancer epigenetics: from mechanism to therapy. Cell 2012;150:12–27. <u>https://doi.org/10.1016/j.cell.2012.06.013</u>.

- [2] Hewezi T. Epigenetic Mechanisms in Nematode-Plant Interactions. Annu Rev Phytopathol 2020;58:119–38. <u>https://doi.org/10.1146/annurev-phyto-010820-012805</u>.
- [3] Marco A, Meharena HS, Dileep V, Raju RM, Davila-Velderrain J, et al. Mapping the epigenomic and transcriptomic interplay during memory formation and recall in the hippocampal engram ensemble. Nat Neurosci 2020;23:1606–17. <u>https://doi.org/10.1038/s41593-020-00717-0</u>.
- [4] Pinello L, Lo Bosco G, Yuan GC. Applications of alignment-free methods in epigenomics. Brief Bioinform 2014;15:419–30. <u>https://doi.org/10.1093/bib/ bbt078</u>.
- [5] Abbott DH. Neuronal androgen receptor: Molecular gateway to polycystic ovary syndrome? Proc Natl Acad Sci U S A 2017;114:4045–7. <u>https://doi.org/ 10.1073/pnas.1703436114</u>.
- [6] Aristizabal MJ, Anreiter I, Halldorsdottir T, Odgers CL, McDade TW, et al. Biological embedding of experience: A primer on epigenetics. Proc Natl Acad Sci U S A 2020;117:23261–9. <u>https://doi.org/10.1073/pnas.1820838116</u>.
- [7] Oleksiewicz U, Machnik M. Causes, effects, and clinical implications of perturbed patterns within the cancer epigenome. Semin Cancer Biol. 2020; S1044-579X(20)30274-1. 10.1016/j.semcancer.2020.12.014.
- [8] Faulk C, Dolinoy DC. Timing is everything: the when and how of environmentally induced changes in the epigenome of animals. Epigenetics 2011;6:791–7. <u>https://doi.org/10.4161/epi.6.7.16209</u>.
- [9] Lozupone M, D'Urso F, Piccininni C, Montagna M, Sardone R, et al. The relationship between epigenetics and microbiota in neuropsychiatric diseases. Epigenomics 2020;12:1559–68. <u>https://doi.org/10.2217/epi-2020-0053</u>.
- [10] O'Rourke CJ, Lafuente-Barquero J, Andersen JB. Epigenome remodeling in cholangiocarcinoma. Trends Cancer 2019;5:335–50. <u>https://doi.org/10.1016/j. trecan.2019.05.002</u>.
- [11] Xia X, Wu WKK, Wong SH, Liu D, Kwong TNY, et al. Bacteria pathogens drive host colonic epithelial cell promoter hypermethylation of tumor suppressor genes in colorectal cancer. Microbiome 2020;8:108. <u>https://doi.org/10.1186/ s40168-020-00847-4</u>.
- [12] Qin Y, Wade PA. Crosstalk between the microbiome and epigenome: messages from bugs. J Biochem 2018;163:105–12. <u>https://doi.org/10.1093/jb/mvx080</u>.
- [13] Alam R, Abdolmaleky HM, Zhou JR. Microbiome, inflammation, epigenetic alterations, and mental diseases. Am J Med Genet B Neuropsychiatr Genet 2017;174:651-60. <u>https://doi.org/10.1002/aimg.b.32567</u>.
- [14] Ansari I, Raddatz G, Gutekunst J, Ridnik M, Cohen D, et al. The microbiota programs DNA methylation to control intestinal homeostasis and inflammation. Nat Microbiol 2020;5:610–9. <u>https://doi.org/10.1038/s41564-019-0659-3</u>.
- [15] Gerhauser C. Impact of dietary gut microbial metabolites on the epigenome. Philos Trans R Soc Lond B Biol Sci 2018;373:20170359. <u>https://doi.org/10.1098/rstb.2017.0359</u>.
- [16] Amatullah H, Jeffrey KL. Epigenome-metabolome-microbiome axis in health and IBD. Curr Opin Microbiol 2020;56:97–108. <u>https://doi.org/10.1016/j. mib.2020.08.005</u>.
- [17] Bhat MI, Kapila R. Dietary metabolites derived from gut microbiota: critical modulators of epigenetic changes in mammals. Nutr Rev 2017;75:374–89. <u>https://doi.org/10.1093/nutrit/nux001</u>.
- [18] D'Aquila P, Carelli LL, De Rango F, Passarino G, Bellizzi D. Gut microbiota as important mediator between diet and DNA methylation and histone modifications in the host. Nutrients 2020;12:597. <u>https://doi.org/10.3390/ nu12030597</u>.
- [19] Silbergleit M, Vasquez AA, Miller CJ, Sun J, Kato I. Oral and intestinal bacterial exotoxins: Potential linked to carcinogenesis. Prog Mol Biol Transl Sci 2020;171:131–93. <u>https://doi.org/10.1016/bs.pmbts.2020.02.004</u>.
- [20] Wu J, Zhao Y, Wang X, Kong L, Johnston LJ, et al. Dietary nutrients shape gut microbes and intestinal mucosa via epigenetic modifications. Crit Rev Food Sci Nutr 2020;1–15. <u>https://doi.org/10.1080/10408398.2020.1828813.</u>
- [21] Krautkramer KA, Kreznar JH, Romano KA, Vivas EI, Barrett-Wilt GA, et al. Diet-Microbiota Interactions Mediate Global Epigenetic Programming in Multiple Host Tissues. Mol Cell 2016;64:982–92. <u>https://doi.org/10.1016/</u> imolcel.2016.10.025.
- [22] Holmes E, Li JV, Marchesi JR, Nicholson JK. Gut microbiota composition and activity in relation to host metabolic phenotype and disease risk. Cell Metab 2012;16:559–64. <u>https://doi.org/10.1016/j.cmet.2012.10.007</u>.
- [23] Chen B, Sun L, Zhang X. Integration of microbiome and epigenome to decipher the pathogenesis of autoimmune diseases. J Autoimmun 2017;83:31–42. <u>https://doi.org/10.1016/j.jaut.2017.03.009</u>.
- [24] Bierne H, Pourpre R. Bacterial Factors Targeting the Nucleus: The Growing Family of Nucleomodulins. Toxins (Basel) 2020;12:220. <u>https://doi.org/</u> 10.3390/toxins12040220.
- [25] Niller HH, Masa R, Venkei A, Meszaros S, Minarovits J. Pathogenic mechanisms of intracellular bacteria. Curr Opin Infect Dis 2017;30:309–15. <u>https://doi.org/ 10.1097/OCO.00000000000363</u>.
- [26] Asrat S, Davis KM, Isberg RR. Modulation of the host innate immune and inflammatory response by translocated bacterial proteins. Cell Microbiol 2015;17:785–95. <u>https://doi.org/10.1111/cmi.12445</u>.
- [27] Botta GA, Arzese A, Minisini R, Trani G. Role of structural and extracellular virulence factors in gram-negative anaerobic bacteria. Clin Infect Dis 1994;18 (Suppl 4):S260–4. <u>https://doi.org/10.1093/clinids/18.supplement 4.s260</u>.
- [28] Popa CM, Tabuchi M, Valls M. Modification of Bacterial Effector Proteins Inside Eukaryotic Host Cells. Front Cell Infect Microbiol 2016;6:73. <u>https://doi.org/</u> 10.3389/fcimb.2016.00073.

- [29] Martins MD, Jiao Y, Larsson L, Almeida LO, Garaicoa-Pazmino C, et al. Epigenetic Modifications of Histones in Periodontal Disease. J Dent Res 2016;95:215–22. <u>https://doi.org/10.1177/0022034515611876</u>.
- [30] Kumar H, Lund R, Laiho A, Lundelin K, Ley RE, et al. Gut microbiota as an epigenetic regulator: pilot study based on whole-genome methylation analysis. mBio 2014;5:e02113-e2114. <u>https://doi.org/10.1128/mBio.02113-14</u>.
- [31] Wang Q, Ye J, Fang D, Lv L, Wu W, et al. Multi-omic profiling reveals associations between the gut mucosal microbiome, the metabolome, and host DNA methylation associated gene expression in patients with colorectal cancer. BMC Microbiol 2020;20:83. <u>https://doi.org/10.1186/s12866-020-01762-2</u>.
- [32] Ye J, Wu W, Li Y, Li L. Influences of the Gut Microbiota on DNA Methylation and Histone Modification. Dig Dis Sci 2017;62:1155–64. <u>https://doi.org/10.1007/ s10620-017-4538-6</u>.
- [33] Yu F, Li K, Li S, Liu J, Zhang Y, et al. CFEA: a cell-free epigenome atlas in human diseases. Nucleic Acids Res 2020;48:D40-4. <u>https://doi.org/10.1093/nar/gkz715</u>.
- [34] Singh Nanda J, Kumar R, Raghava GP. dbEM: A database of epigenetic modifiers curated from cancerous and normal genomes. Sci Rep 2016;6:19340. <u>https:// doi.org/10.1038/srep19340</u>.
- [35] Turinsky AL, Turner B, Borja RC, Gleeson JA, Heath M, et al. DAnCER: diseaseannotated chromatin epigenetics resource. Nucleic Acids Res 2011;39: D889–94. <u>https://doi.org/10.1093/nar/gkq857</u>.
- [36] Liu D, Zhao L, Wang Z, Zhou X, Fan X, et al. EWASdb: epigenome-wide association study database. Nucleic Acids Res 2019;47:D989–93. <u>https://doi.org/10.1093/nar/gkv942</u>.
- [37] Ding W, Chen J, Feng G, Chen G, Wu J, et al. DNMIVD: DNA methylation interactive visualization database. Nucleic Acids Res 2020;48:D856–62. https://doi.org/10.1093/nar/gkz830.
- [38] Xiong Z, Li M, Yang F, Ma Y, Sang J, et al. EWAS Data Hub: a resource of DNA methylation array data and metadata. Nucleic Acids Res 2020;48:D890–5. <u>https://doi.org/10.1093/nar/gkz840</u>.
- [39] Deng G, Yang J, Zhang Q, Xiao ZX, Cai H. MethCNA: a database for integrating genomic and epigenomic data in human cancer. BMC Genomics 2018;19:138. https://doi.org/10.1186/s12864-018-4525-0.
- [40] Xie B, Yuan Z, Yang Y, Sun Z, Zhou S, et al. MOBCdb: a comprehensive database integrating multi-omics data on breast cancer for precision medicine. Breast Cancer Res Treat 2018;169:625–32. <u>https://doi.org/10.1007/s10549-018-4708-z</u>.
- [41] Xin Y, Chanrion B, O'Donnell AH, Milekic M, Costa R, et al. MethylomeDB: a database of DNA methylation profiles of the brain. Nucleic Acids Res 2012;40: D1245–9. <u>https://doi.org/10.1093/nar/gkr1193</u>.
- [42] Xiong Y, Wei Y, Gu Y, Zhang S, Lyu J, et al. DiseaseMeth version 2.0: a major expansion and update of the human disease methylation database. Nucleic Acids Res 2017;45:D888–95. <u>https://doi.org/10.1093/nar/gkw1123</u>.
- [43] Li M, Zou D, Li Z, Gao R, Sang J, et al. EWAS Atlas: a curated knowledgebase of epigenome-wide association studies. Nucleic Acids Res 2019;47:D983–8. https://doi.org/10.1093/nar/gky1027.
- [44] Roberts RJ, Vincze T, Posfai J, Macelis D. REBASE-a database for DNA restriction and modification: enzymes, genes and genomes. Nucleic Acids Res 2015;43: D298-9. <u>https://doi.org/10.1093/nar/gku1046</u>.
- [45] Amoreira C, Hindermann W, Grunau C. An improved version of the DNA Methylation database (MethDB). Nucleic Acids Res 2003;31:75–7. <u>https://doi.org/10.1093/nar/gkg093</u>.
- [46] Sood AJ, Viner C, Hoffman MM. DNAmod: the DNA modification database. J Cheminform 2019;11:30. <u>https://doi.org/10.1186/s13321-019-0349-4</u>.
- [47] Xuan Lin QX, Sian S, An O, Thieffry D, Jha S, et al. MethMotif: an integrative cell specific database of transcription factor binding motifs coupled with DNA methylation profiles. Nucleic Acids Res 2019;47:D145–54. <u>https://doi.org/ 10.1093/nar/gkv1005</u>.
- [48] Zhang Y, Lv J, Liu H, Zhu J, Su J, et al. HHMD: the human histone modification database. Nucleic Acids Res 2010;38:D149–54. <u>https://doi.org/10.1093/nar/gkp968</u>.
- [49] Choi J, Kim KT, Huh A, Kwon S, Hong C et al. dbHiMo: a web-based epigenomics platform for histone-modifying enzymes. Database (Oxford). 2015;2015:bav052. 10.1093/database/bav052.
- [50] Sharma G, Sowpati DT, Singh P, Khan MZ, Ganji R et al. Genome-wide non-CpG methylation of the host genome during M. tuberculosis infection. Sci Rep. 2016;6:25006. 10.1038/srep25006
- [51] Atlante S, Mongelli A, Barbi V, Martelli F, Farsetti A et al. The epigenetic implication in coronavirus infection and therapy. Clin Epigenet 12:156. 10.1186/s13148-020-00946-x
- [52] Sayers EW, Beck J, Bolton EE, Bourexis D, Brister JR, et al. Database resources of the National Center for Biotechnology Information. Nucleic Acids Res 2021;49: D10–7. <u>https://doi.org/10.1093/nar/gkaa892</u>.
- [53] Li R, Huang X, Liang X, Su M, Lai KP, et al. Integrated omics analysis reveals the alteration of gut microbe-metabolites in obese adults. Brief Bioinform 2017;18:98–104. <u>https://doi.org/10.1093/bib/bbaa165</u>.
- [54] Marino E, Richards JL, McLeod KH, Stanley D, Yap YA, et al. Gut microbial metabolites limit the frequency of autoimmune T cells and protect against type 1 diabetes. Nat Immunol 2017;18(11):1271. <u>https://doi.org/10.1038/</u> ni1117-1271c.

- [55] Elhag DA, Kumar M, Al KS. Exploring the triple interaction between the host genome, the epigenome, and the gut microbiome in type 1 diabetes. Int J Mol Sci 2020;22:125. <u>https://doi.org/10.3390/ijms22010125</u>.
- [56] Ragusa M, Santagati M, Mirabella F, Lauretta G, Cirnigliaro M, et al. Potential associations among alteration of salivary miRNAs, saliva microbiome structure, and cognitive impairments in autistic children. I Int J Mol Sci 2020;21:6203. <u>https://doi.org/10.3390/ijms21176203</u>.
- [57] Popov J, Bandura J, Markovic F, Borojevic R, Anipindi VC et al. Influence of bacterial components on the developmental programming of enteric neurons. Physiol Rep. 2020;8:e14611. 10.14814/phy2.14611
- [58] Dhalech AH, Fuller TD, Robinson CM. Specific bacterial cell wall components influence the stability of coxsackievirus B3. J Virol 2021;95(22):e0142421.
- [59] Sharma G, Upadhyay S, Srilalitha M, Nandicoori VK, Khosla S. The interaction of mycobacterial protein Rv2966c with host chromatin is mediated through non-CpG methylation and histone H3/H4 binding. Nucleic Acids Res 2015;43:3922–37. <u>https://doi.org/10.1093/nar/gkv261</u>.
- [60] Yaseen I, Kaur P, Nandicoori VK, Khosla S. Mycobacteria modulate host epigenetic machinery by Rv1988 methylation of a non-tail arginine of histone H3. Nat Commun. 2015;6:8922. ARTN892210.1038/ncomms9922
- [61] Zheng L, Leung ETY, Wong HK, Lui G, Lee N, et al. Unraveling methylation changes of host macrophages in Mycobacterium tuberculosis infection. Tuberculosis 2016;98:139–48. <u>https://doi.org/10.1016/j.tube.2016.03.003</u>.
- [62] Pacis A, Tailleux L, Morin AM, Lambourne J, MacIsaac JL, et al. Bacterial infection remodels the DNA methylation landscape of human dendritic cells. Genome Res 2015;25:1801–11. <u>https://doi.org/10.1101/gr.192005.115</u>.
- [63] Zhu YL, Jiang QL, Lou XJ, Ji XW, Wen ZZ, et al. MicroRNAs up-regulated by CagA of helicobacter pylori induce intestinal metaplasia of gastric epithelial cells. PLoS ONE 2012;7:e35147.
- [64] Federhen S. Type material in the NCBI taxonomy database. Nucleic Acids Res 2015;43:D1086–98. <u>https://doi.org/10.1093/nar/gku1127</u>.
- [65] Tang J, Wu X, Mou M, Wang C, Wang L, et al. GIMICA: host genetic and immune factors shaping human microbiota. Nucleic Acids Res 2021;49:D715-22. https://doi.org/10.1093/nar/gkaa851.
- [66] Cheng L, Qi C, Zhuang H, Fu T, Zhang X. gutMDisorder: a comprehensive database for dysbiosis of the gut microbiota in disorders and interventions. Nucleic Acids Res 2020;48:D554–60. <u>https://doi.org/10.1093/nar/gkz843</u>.
- [67] Noronha A, Modamio J, Jarosz Y, Guerard E, Sompairac N, et al. The Virtual Metabolic Human database: integrating human and gut microbiome metabolism with nutrition and disease. Nucleic Acids Res 2019;47:D614–24. https://doi.org/10.1093/nar/gkv992.
- [68] UniProt C. UniProt: a worldwide hub of protein knowledge. Nucleic Acids Res 2019;47:D506-15. <u>https://doi.org/10.1093/nar/gky1049</u>.
- [69] Zhao WY, Wang YF, Liang FS. Chemical and light inducible epigenome editing. Int J Mol Sci 2020;21(3):998. <u>https://doi.org/10.3390/ijms21030998</u>.
- [70] Ko YA, Susztak K. Epigenomics: the science of no-longer-junk DNA. Why study it in chronic kidney disease? Semin Nephrol 2013;33:354–62. <u>https://doi.org/ 10.1016/j.semnephrol.2013.05.007</u>.
- [71] Polak P, Karlic R, Koren A, Thurman R, Sandstrom R, et al. Cell-of-origin chromatin organization shapes the mutational landscape of cancer. Nature 2015;518:360–4. <u>https://doi.org/10.1038/nature14221</u>.
- [72] Robertson AB, Dahl JA, Vagbo CB, Tripathi P, Krokan HE, et al. A novel method for the efficient and selective identification of 5-hydroxymethylcytosine in genomic DNA. Nucleic Acids Res 2011;39(8):e55.
- [73] Teng CS, Wu BH, Yen MR, Chen PY. MethGET: web-based bioinformatics software for correlating genome-wide DNA methylation and gene expression. BMC Genomics 2020;21(1):375. <u>https://doi.org/10.1186/s12864-020-6722-x</u>.
- [74] Deng SL, Chua NH. Inverted-repeat RNAs targeting FT intronic regions promote FT expression in arabidopsis. Plant Cell Physiol 2015;56(8):1667–78. <u>https:// doi.org/10.1093/pcp/pcv091</u>.
- [75] Yates AD, Achuthan P, Akanni W, Allen J, Allen J, et al. Ensembl 2020. Nucleic Acids Res 2020;48:D682–8. <u>https://doi.org/10.1093/nar/gkz966</u>.
- [76] Mukherjee S, Stamatis D, Bertsch J, Ovchinnikova G, Katta HY, et al. Genomes OnLine database (GOLD) vol 7: updates and new features. Nucleic Acids Res 2019;47:D649–59. <u>https://doi.org/10.1093/nar/gky977</u>.
- [77] Wishart DS, Feunang YD, Marcu A, Guo AC, Liang K, et al. HMDB 4.0: the human metabolome database for 2018. Nucleic Acids Res 2018;46:D608–17. https://doi.org/10.1093/nar/gkx1089.
- [78] Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K. KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic Acids Res 2017;45:D353–61. <u>https://doi.org/10.1093/nar/gkw1092</u>.
- [79] Smith CA, O'aille G, Want EJ, Qin C, Trauger SA, et al. METLIN: a metabolite mass spectral database. Ther Drug Monit 2005;27:747–51. <u>https://doi.org/ 10.1097/01.ftd.0000179845.53213.39.</u>
- [80] The L. Icd-11. Lancet 2019;393:2275. <u>https://doi.org/10.1016/S0140-6736(19)</u> 31205-X.
- [81] Hamon MA, Cossart P. Histone modifications and chromatin remodeling during bacterial infections. Cell Host Microbe 2008;4:100–9. <u>https://doi.org/ 10.1016/j.chom.2008.07.009</u>.
- [82] Stols-Goncalves D, Tristao LS, Henneman P, Nieuwdorp M. Epigenetic markers and microbiota/metabolite-induced epigenetic modifications in the pathogenesis of obesity, metabolic syndrome, type 2 diabetes, and nonalcoholic fatty liver disease. Curr Diab Rep 2019;19(6):31. <u>https://doi.org/ 10.1007/s11892-019-1151-4</u>.

L. Wang, W. Zhang, X. Wu et al.

Computational and Structural Biotechnology Journal 20 (2022) 2455-2463

- [83] Hennessy EJ, FitzGerald GA. Battle for supremacy: nucleic acid interactions between viruses and cells. J Clin Invest 2021;131(3):e144227.
 [84] Rizzo HE, Escaname EN, Alana NB, Lavender E, Gelfond J, et al. Maternal
- 84] Rizzo HE, Escaname EN, Alana NB, Lavender E, Gelfond J, et al. Maternal diabetes and obesity influence the fetal epigenome in a largely Hispanic population. Clin Epigenetics 2020;12(1):34. <u>https://doi.org/10.1186/s13148-020-0824-9</u>.
- [85] Rehan VK, Liu J, Naeem E, Tian J, Sakurai R, et al. Perinatal nicotine exposure induces asthma in second generation offspring. Bmc Med 2012;10:129. <u>https://doi.org/10.1186/1741-7015-10-129</u>.
- [86] Shamsi MB, Firoz AS, Imam SN, Alzaman N, Samman MA. Epigenetics of human diseases and scope in future therapeutics. J Taibah Univ Med Sci 2017;12:205–11. <u>https://doi.org/10.1016/j.jtumed.2017.04.003</u>.